# Mutational Signatures in Cytogenetic Risk Groups of De Novo AML and MDS

Robyn Sussman, PhD
Center for Personalized Diagnostics
AMP webinar: 10/4/18



### Myeloid neoplasms

- Blast count
  - MDS/MPN<20%<AML
- Genetic features
  - FISH
  - Cytogenetics
  - Sequencing
- Displasia
  - (MDS)

### Other myeloproliferative neoplasms (MPN)

- BCR-ABL (Ph+)
  - Chronic Myelogenous Leukemia (CML)
- Ph-
  - Polycythemia vera (PV)
  - Essential thrombocytopenia (AT)
  - Primary myelofibrosis (PMF)

### Myeloid neoplasms are heterogeneous

- Cytogenetic abnormalities are associated with prognosis
- Many subtypes exist with multiple overlapping mutations
- Recurrent mutations belong to several distinct pathways
- Pre-leukemic and leukemic cells undergo clonal evolution
  - Heterogeneous cell populations with mutations conferring different functional properties

### Genetic basis of myeloid neoplasms

- De novo AML
  - NPM1, CBF and KMT2A mutations
- MDS
  - Progression to AML assacciated with mutations in TP53, RUNX1, ETV6, EZH2, ASXL1
- sAML
  - Spliceosome complex: SRSF2, SF3B1, U2AF1, ZRSR2
  - Epigenetic regulators: ASXL1, EZH2, BCOR
  - Cohesion Complex: STAG2
  - Many mutations from MDS or myelofibrosis and are retained after transformation

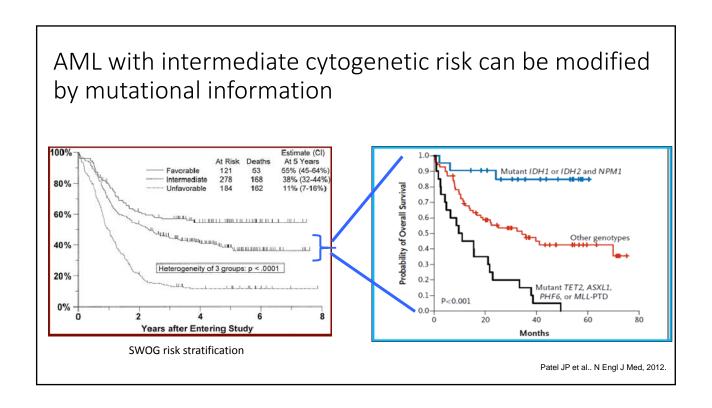
# AML risk stratification: Medical Research Council (MRC) & Southwest Oncology Group (SWOG)

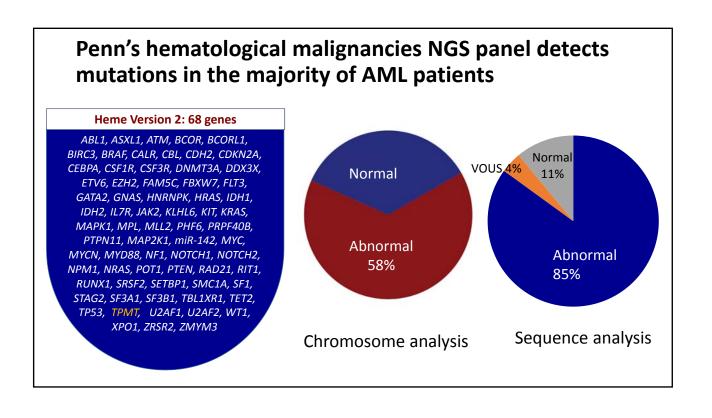
Risk Status	SWOG[1]	CALGB[2] <sup>a</sup>	MRC (1998)[3]	MRC (2010)[4]
Favorable	t(15;17), t(8;21), inv(16)/t(16;16)/ del(16q)	(8;21), inv(16)/t(16;16)	t(15;17), t(8;21), inv(16)/t(16;16)/ del(16q)	t(15;17)(q22;q21), t(8;21) (q22;q22), inv(16) (p13q22)/t(16;16)(p13;q22)
Intermediate	Normal, +8, +6, -Y, del(12p)	Normal, –Y, del(5q), t(6;9), t(6;11), –7, loss of 7q, +8 sole, +8 with 1 other abnormality, del(9q), t(9;11), +11, del(11q), t(11;19)(q23;p13.1), +13, del(20q), +21	Normal, 11q23 abn, +8, del(9q), del(7q), +21, +22, all others	Abnormalities not classified as favorable or unfavorable
Unfavorable	abn(3q), del(5q)/ –5, –7/del(7q), t(6;9), t(9;22), 9q, 11q, 20q, 21q, 17p, complex (≥ 3 unrelated abnormalities)	inv(3)/t(3,3), abn(12p), complex (2 3 unrelated abnormalities)	abn(3q), del(5q)/ −5, −7, complex (≥ 5 unrelated abnormalities)	abn(3q) [excluding t(3;5) (q21–25;q31–35)], inv(3) (q21–26;q41–35), inv(3) (q21q26)/t(3;3)(q21;q26), add(5q), del(5q), –5, add (7q)/del(7q), –7, t(6;11) (q27;q23), t(10;11) (p11–13;q23), t(11;q23) [excluding t(9;11) (p21–22;q23) and t(11;19) (q23;p13)],t(9;22)(q34;q11), –17/ abn(17p),complex (2 4 unrelated abnormalities)
Unknown	All other abnormalities	Category not recognized	Category not recognized	Category not recognized

Orozco, et.al., 2012

### AML risk stratification: European LeukemiaNet (ELN)

Risk category*	Genetic abnormality			
Favorable	t(8;21)(q22;q22.1); RUNXI-RUNXITI			
	inv(16)(p13.1q22) or t(16;16)(p13.1;q22); CBFB-MYH11			
	Mutated NPM1 without FLT3-ITD or with FLT3-ITD low†			
	Biallelic mutated CEBPA			
Intermediate	Mutated NPM1 and FLT3-ITD high†			
	Wild-type NPMI without FLT3-ITD or with FLT3-ITD low† (without ad-	verse-risk genetic lesions)		
	t(9;11)(p21.3;q23.3); MLLT3-KMT2A <sup>†</sup>			
	Cytogenetic abnormalities not classified as favorable or adverse			
Adverse	t(6,9)(p23;q34.1); DEK-NUP214 Frequencies, response rates, and outcome measures should be reported by ri available, by specific genetic lesions indicated.	rates, and outcome measures should be reported by risk category, and, if sufficient numbers ar		
	t(v;11q23.3); KMT2A rearranged		of a marker is treatment-dependent and may change with new therapies.	
	t(9;22)(q34.1;q11.2); BCR-ABL1		(<0.5); high, high allelic ratio (≥0.5); semiquantitative assessment of FLT3-ITD allelic ratio	
	inv(3)(q21.3q26.2) or t(3;3)(q21.3;q26.2); GATA2,MECOM(EVT1)		nt analysis) is determined as ratio of the area under the curve "FLT3-ITD" divided by area under	
	-5 or del(5q); -7; -17/abn(17p)		ype"; recent studies indicate that AML with $NPMI$ mutation and $FLT3$ -ITD low allelic ratio avorable prognosis and patients should not routinely be assigned to allogeneic $HCT_{}^{.5159,.77}$	
	Complex karyotype, § monosomal karyotype	The presence of t(9;11)(p21.3;q23.3) takes precedence over rare, concurrent adverse-risk gene mutations.  Three or more unrelated chromosome abnormalities in the absence of 1 of the WHO-designated recurring translocations or inversions, that is, t8.2.1), inv(16) or t1(6.16), t9.111, tv.111/vv.23.3, t(6.9.6), inv(3) or t(3.3).		
	Wild-type NPMI and FLT3-ITD high†			
	Mutated RUNXI	with BCR-ABL1.		
	Mutated ASXL1		ce of 1 single monosomy (excluding loss of X or Y) in association with at least 1 additional 1 chromosome abnormality (excluding core-binding factor AML). 116	
	Mutated TP53#	These markers should not be used as an adverse prognostic marker if they co-occur with favorable-risk AML		
		subtypes. #TP53 mutations are si	gnificantly associated with AML with complex and monosomal karyotype. 37,66-69	
			Döhner et.al., 2017	





### **Evolution of the heme panel**

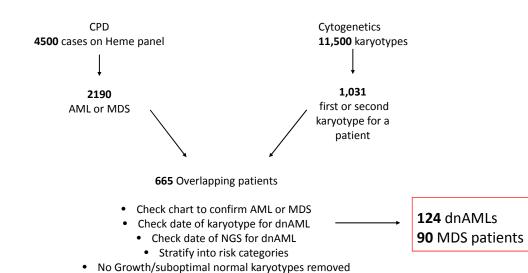
#### Heme Version 1: 33 genes

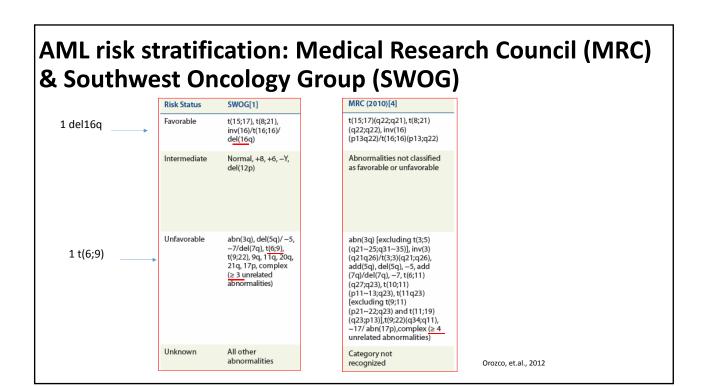
ASXL1, ATM, BRAF, CBL, CDKN2A, DDX3X, DNMT3A, ETV6, EZH2, FBXW7, FLT3, GNAS, IDH1, IDH2, JAK2, KLHL6, KIT, KRAS, MAPK1, PHF6, PTPN11, MYD88, NOTCH1, NPM1, NRAS, PTEN, RUNX1, SF3B1, TET2, TP53, WT1, XPO1, ZMYM3

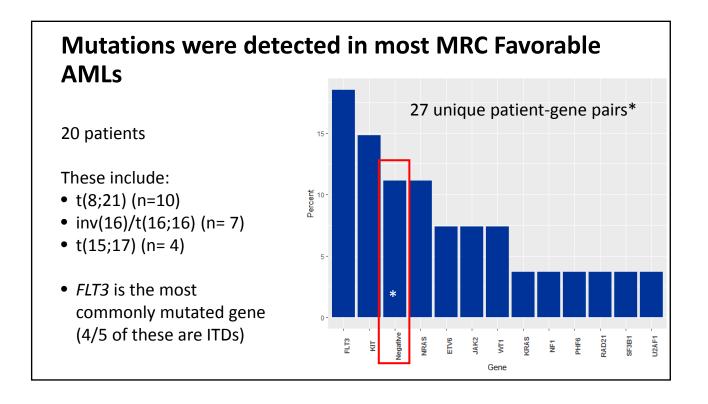
#### Heme Version 2: 68 genes

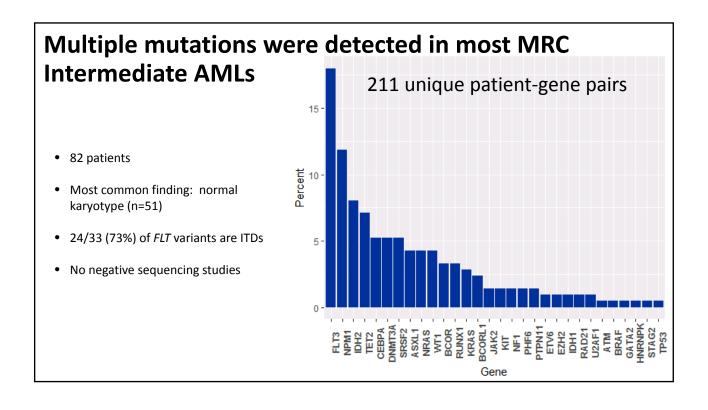
ABL1, ASXL1, ATM, BCOR, BCORL1, BIRC3, BRAF, CALR, CBL, CDH2, CDKN2A, CEBPA, CSF1R, CSF3R, DNMT3A, DDX3X, ETV6, EZH2, FAM5C, FBXW7, FLT3, GATA2, GNAS, HNRNPK, HRAS, IDH1, IDH2, IL7R, JAK2, KLHL6, KIT, KRAS, MAPK1, MPL, MLL2, PHF6, PRPF40B, PTPN11, MAP2K1, miR-142, MYC, MYCN, MYD88, NF1, NOTCH1, NOTCH2, NPM1, NRAS, POT1, PTEN, RAD21, RIT1, RUNX1, SRSF2, SETBP1, SMC1A, SF1, STAG2, SF3A1, SF3B1, TBL1XR1, TET2, TP53, TPMT, U2AF1, U2AF2, WT1, XPO1, ZRSR2, ZMYM3

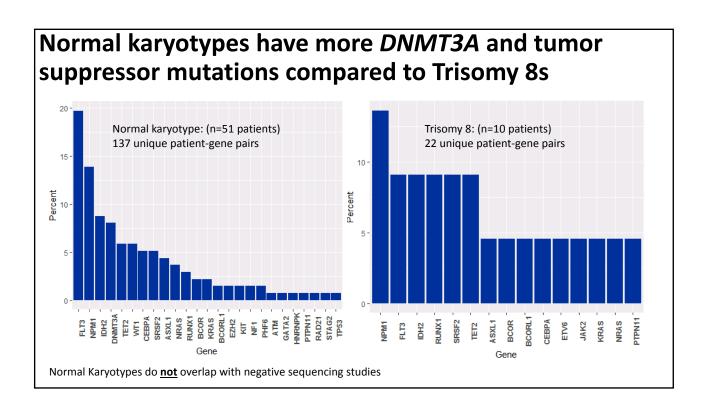
### **Overlapping Risk Stratification with Mutations** from CPD

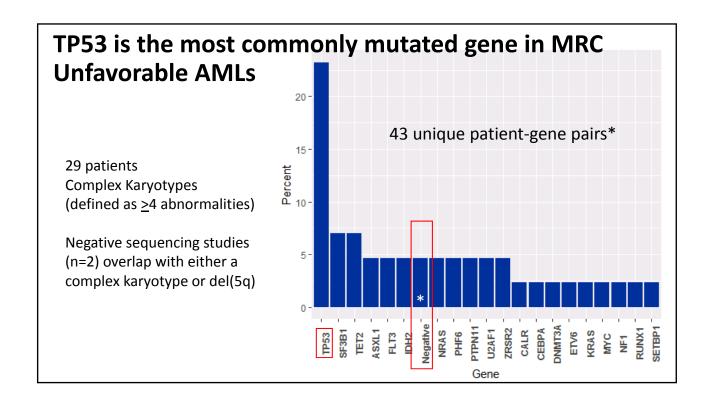


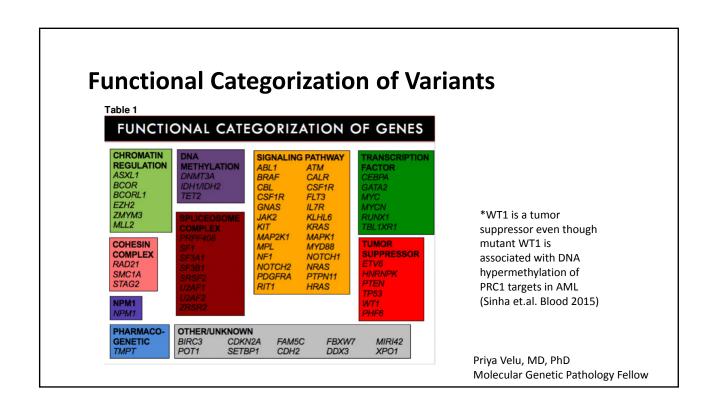


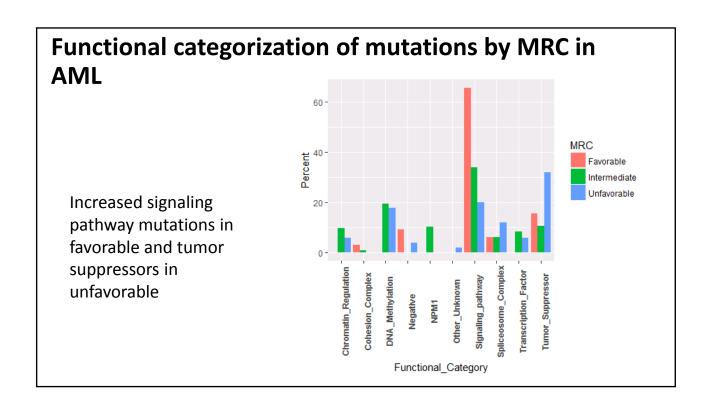


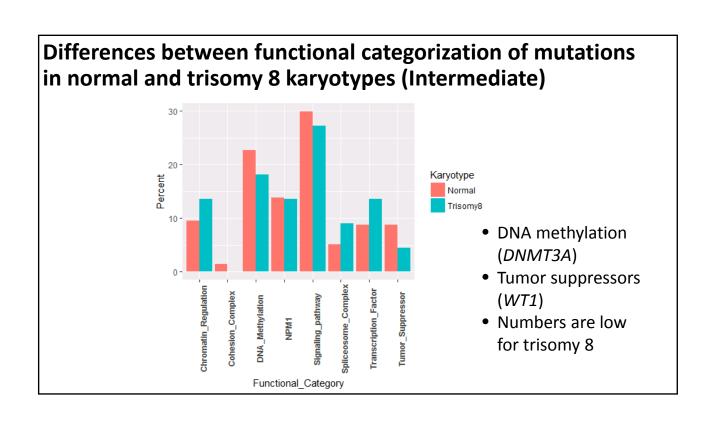












### Main conclusions from AML (MRC)

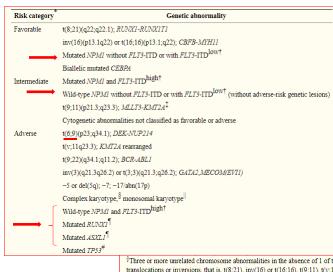
- "Negative" sequencing studies exist in both favorable and unfavorable cytogenetic categories. (Limitations of a targeted panel)
- Signaling pathway genes are the most commonly mutated in favorable
  - More variants in favorable than other karyotypes
- No DNA methylation gene mutations in favorable karyotypes
- Tumor Suppressors are mainly mutated in unfavorable karyotypes
  - Almost exclusively TP53
- NPM1 mutations only occur in intermediate karyotypes
- WT1 mutations are common in favorable and intermediate, do not occur in unfavorable
- Within intermediate karyotypes:
  - Tumor suppressors, signaling pathway and DNA methylation genes mutated in more normal karyotypes than trisomy 8s
  - Chromatin regulatory, spliceosome complex and transcription factor genes mutated more in Trisomy 8 than normal karyotypes.

### AML risk stratification: European LeukemiaNet (ELN)

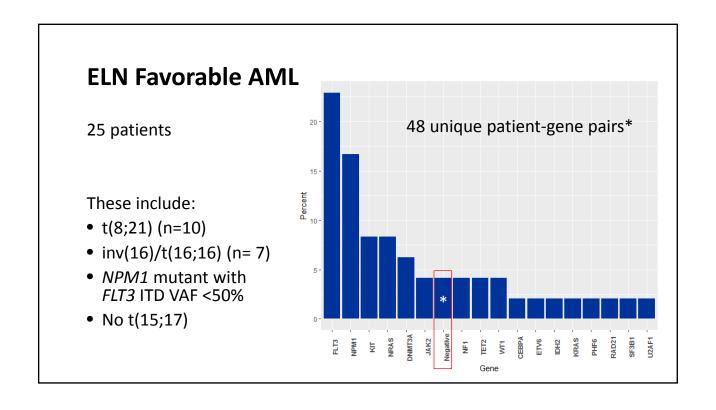
Risk category*	Genetic abnormality		
Favorable	t(8;21)(q22;q22.1); RUNXI-RUNXITI		
	inv(16)(p13.1q22) or t(16;16)(p13.1;q22); CBFB-MYH11		
	Mutated NPM1 without FLT3-ITD or with FLT3-ITD low†		
	Biallelic mutated CEBPA		
intermediate	Mutated NPM1 and FLT3-ITD high†		
	Wild-type NPMI without FLT3-ITD or with FLT3-ITD low† (without adverse-risk genetic lesions)		
	t(9;11)(p21.3;q23.3); MLLT3-KMT2A <sup>‡</sup>		
	Cytogenetic abnormalities not classified as favorable or adverse		
	t(6;9)(p23;q34.1); DEK-NUP214		rates, and outcome measures should be reported by risk category, and, if sufficient numbers a genetic lesions indicated.
	t(v;11q23.3); KMT2A rearranged		t of a marker is treatment-dependent and may change with new therapies.
	t(9;22)(q34.1;q11.2); BCR-ABL1	†Low, low allelic ratio (using DNA fragment a the curve "FLT3-wild t may also have a more f ‡The presence of t(9;11	a manch is dedunctive pendent and may change with new declarges. $(<0.5)$ ; high, high allelic ratio $(\ge0.5)$ ; semiquantitative assessment of FLT3-ITD allelic ratio
	inv(3)(q21.3q26.2) or t(3;3)(q21.3;q26.2); GATA2,MECOM(EVI1)		ent analysis) is determined as ratio of the area under the curve "FLT3-ITD" divided by area under ild type"; recent studies indicate that AML with NPMI mutation and FLT3-ITD low allelic ratio
	-5 or del(5q); -7; -17/abn(17p)		raype , recent studies indicate that AML with MPMI mutation and PLI3-11D low alteric ratio favorable prognosis and patients should not routinely be assigned to allogeneic HCT.57-59.77
	Complex karyotype, <sup>§</sup> monosomal karyotype <sup>∥</sup>		11)(p21.3;q23.3) takes precedence over rare, concurrent adverse-risk gene mutations.
	Wild-type NPMI and FLT3-ITD high†		ted chromosome abnormalities in the absence of 1 of the WHO-designated recurring rsions, that is, t(8:21), inv(16) or t(16:16), t(9:11), t(v:11)(v:q23.3), t(6:9), inv(3) or t(3:3); AN
	Mutated RUNXI	with BCR-ABL1.	
	Mutated ASXL1		nce of 1 single monosomy (excluding loss of X or Y) in association with at least 1 additional ral chromosome abnormality (excluding core-binding factor AML). 116
	Mutated TP53 <sup>#</sup>	<sup>¶</sup> These markers shoul	d not be used as an adverse prognostic marker if they co-occur with favorable-risk AML
		subtypes. #TP53 mutations are	significantly associated with AML with complex and monosomal karyotype. 37,66-69
			Döhner et.al., 2017

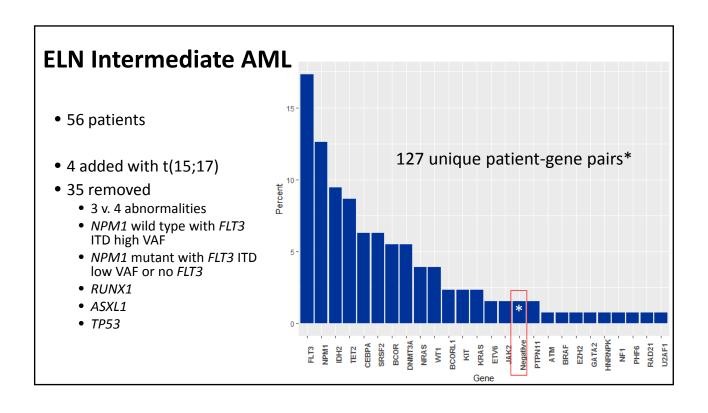
### There are significant differences between MRC and ELN categorization schemes

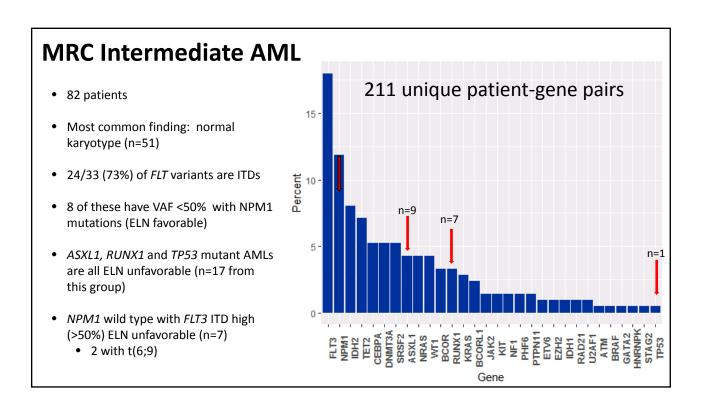


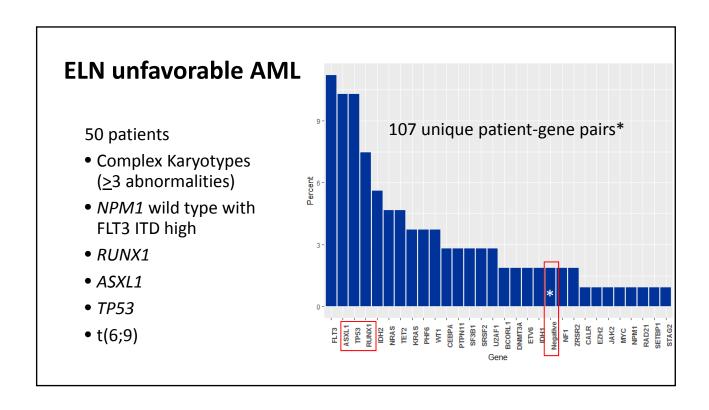


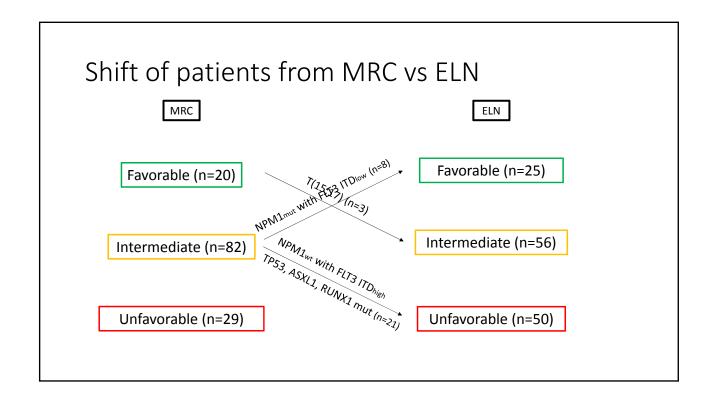
§Three or more unrelated chromosome abnormalities in the absence of 1 of the WHO-designated recurring translocations or inversions, that is, t(8;21), inv(16) or t(16;16), t(9;11), t(v;11)(v;q23.3), t(6;9), inv(3) or t(3;3); AML with BCR-ABL1.

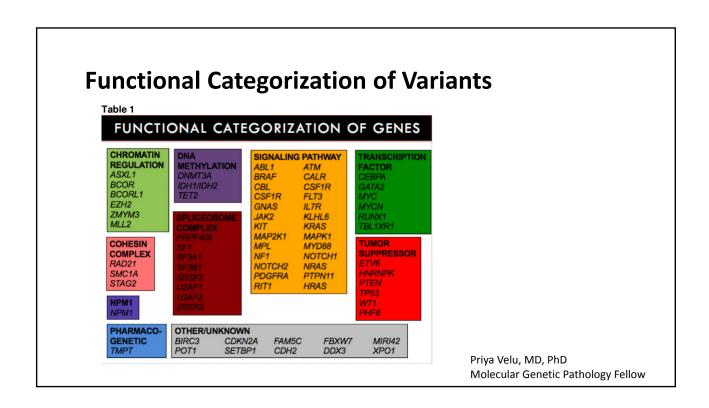


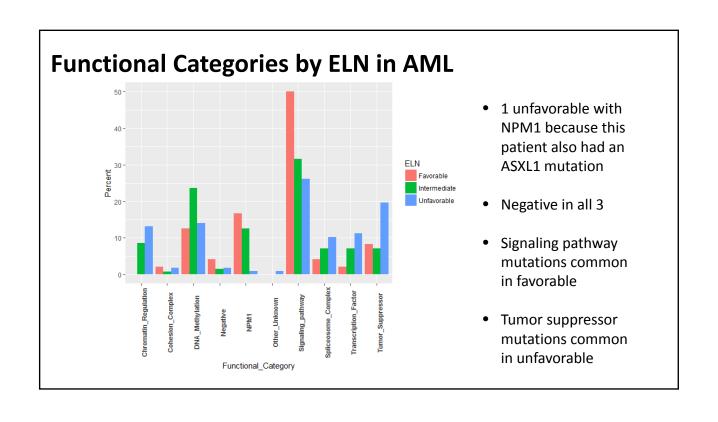


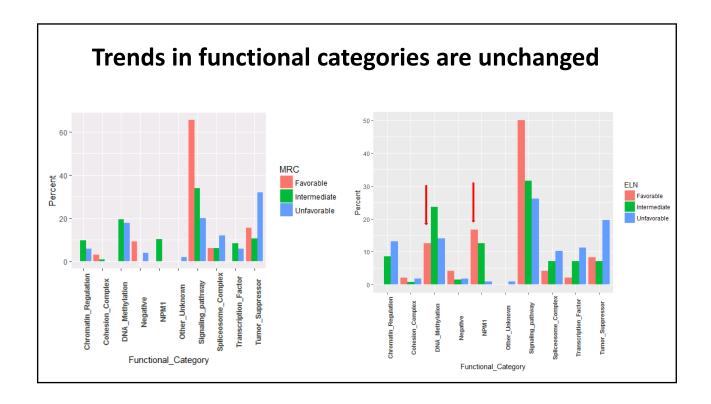










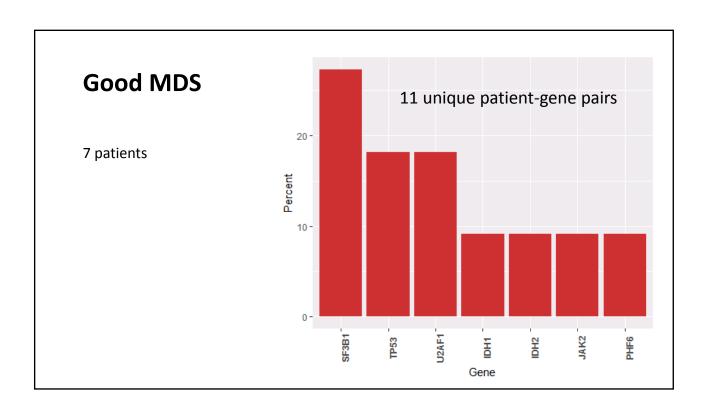


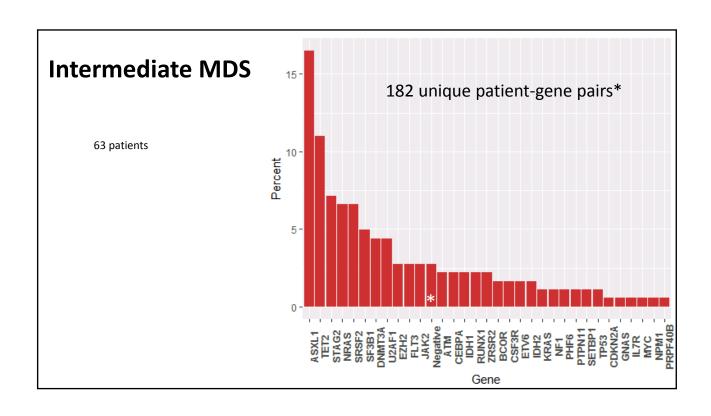
#### Differences between MRC, ELN and SWOG

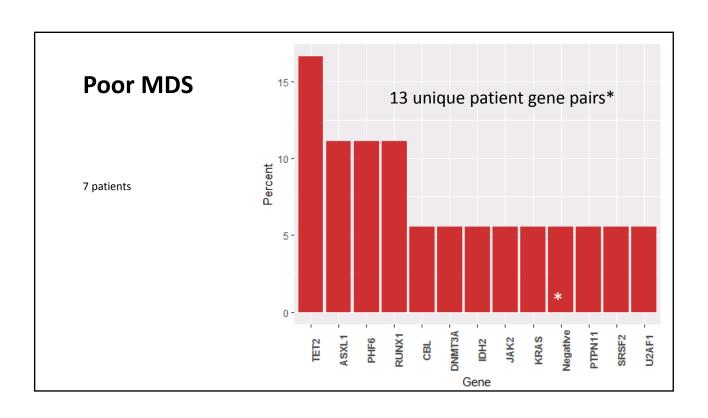
- MRC and SWOG are very similar. Two intermediate cases would be changed (1 to SWOG favorable, 1 to SWOG unfavorable)
  - Therefore no additional analysis with SWOG
- ELN incorporated t(6;9) into unfavorable which moved two cases from intermediate to unfavorable (also had unfavorable mutations)
- ELN favorable does not include t(15;17) in criteria
- ELN includes mutation detection: all ASXL1, RUNX1 and TP53 mutant AMLs are unfavorable
  - NPM1 wild type or mutant with FLT3 ITD
    - FLT3 ITD high = >50% VAF. Need to consider % blasts in the sample
- ELN has more unfavorable AMLs from our cohort

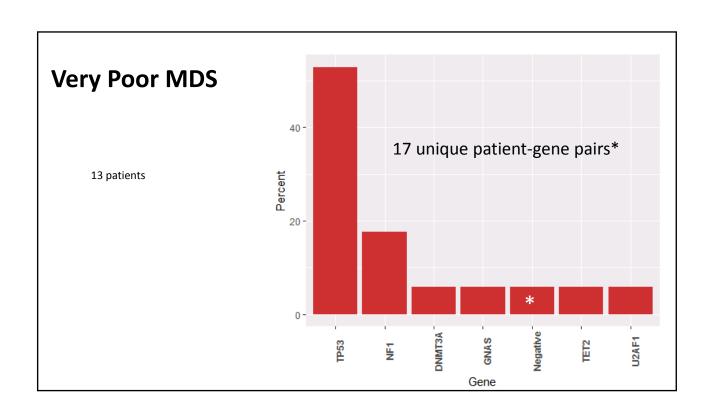
# MDS risk stratification by International Prognostic Scoring System (IPSS-R)

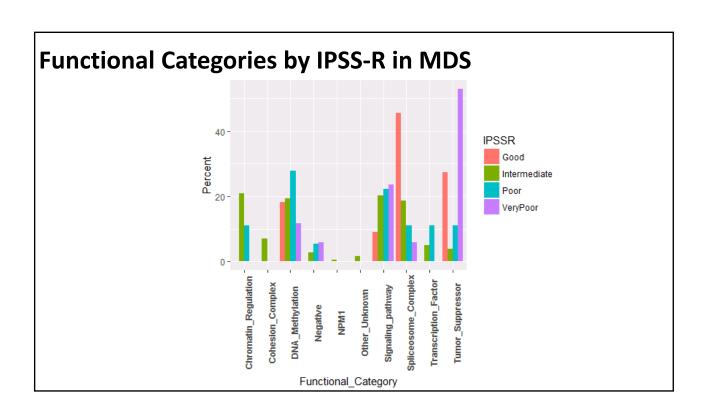
Cytogenetic prognostic subgroups	Cytogenetic abnormalities	
Very good No	Cases with matching sequencing -Y, del(11q)	
Good	Normal, del(5q), del(12p), del(20q), double including del(5q)	
Intermediate	del(7q), +8, +19, i(17q), any other single or double independent clones	
Poor	-7, inv(3)/t(3q)/del(3q), double including -7/del(7q), Complex: 3 abnormalities	
Very poor	Complex: >3 abnormalities	











#### Main conclusions from MDS

- "Negative" sequencing studies exist in 3/4 cytogenetic categories. (Limitations of a targeted panel)
- Spliceosome complex genes are the most commonly mutated in good MDS
  - More variants in good than other karyotypes
- Cohesion complex gene mutations are exclusive to intermediate MDS
- Tumor suppressors are mainly mutated in very poor karyotypes
  - Almost exclusively TP53
- Very few NPM1 mutations in MDS
- The n is low for all risk categories except intermediate

### Overall considerations for study design

- Clean data set with dnAML and new MDS.
  - Could include sAMLs
  - Older MDS
    - For both, karyotypes could be different than the current data set but you could still link risk/prognosis from karyotype to mutational signatures
- WT1 is a tumor suppressor
  - Mutant WT1 can cause a hyper-methylated phenotype in AML
  - This is causal and not a direct function by WT1
- Mutations by functional category trends do not differ between MRC and ELN AML
  - Survival data may show one to be superior to the other
- Use type of alteration as a way to stratify cytogenetics
  - Trisomys, monosomys, translocations, deletions, etc.

### Thank you key players

- Priya Velu, Penn MGP fellow
- Jennifer Morrissette, Director of Cytogenetics and Clinical Director of CPD
- Dan Ackerman, Staff Scientist
- Ashkan Bigdeli, Bioinformatics Specialist
- Beckman Coulter Life Sciences (Genomic Reagents)
  - We have transitioned the majority of our extractions onto FormaPure Total for FFPE, cytology, fresh tissue and are validating for bone cores